

## Antimicrobial Activity of Isolated Soil Fungi

Han Wai Zin<sup>1</sup>, Yee Yee Thu<sup>2</sup>

### Abstract

In this study, the two soil samples were collected from the two different depths (20 cm and 40 cm) near Bago River in (94) Ward, Dagon Seikkan Township, Yangon Region. The twelve soil fungi were isolated from the collected samples on three different media by the serial dilution method. Isolated strains were *Cephalosporium* sp., *Aspergillus* spp., *Penicillium* spp., *Cladosporium* sp., *Madurella* sp., *Paecilomyces* sp., *Hormiscium* sp. and *Gonatotryps* sp. Antimicrobial activity of the fermented broths from all isolated strains was examined on five test organisms by paper disc diffusion method and fermentation was carried out from 4 to 10 days. The fermented broths of all isolated strains (HAN 1 to HAN 12) showed antimicrobial activity on *Agrobacterium tumefaciens* and *Malassezia furfur* from fourth day to ten day fermentations. Crude metabolites of twelve active strains were extracted with ethyl acetate, and their antimicrobial activity was also examined on five test organisms. The crude extracts of twelve strains (HAN 1 to HAN 12) indicated excellent antimicrobial activity on *Agrobacterium tumefaciens*, *Candida albicans*, *Escherichia coli* and *Malassezia furfur* at day 7 fermentation. The most active strains should be chosen to produce the bioactive compounds to protect *Agrobacterium tumefaciens* causing crown gall disease on plants, *Candida albicans* causing alimentary tract and vaginal infections, *Escherichia coli* causing urinary tract infection and *Malassezia furfur* causing dandruff and skin infections on humans.

**Keywords:** Antimicrobial activity, Fermented broth, Soil fungi

### Introduction

Soil microorganisms are classified into bacteria, actinomycetes, fungi, algae and protozoa. Each of these groups has characteristic functions in soils (Mendes *et al.*, 2013). Fungi benefit most plants by suppressing plant root diseases and fungi promote healthier plants by attacking plant pathogens with fungal enzymes (Lowenfels & Lewis, 2006; Sylvia *et al.*, 2005).

The various species of *Penicillium* can colonise many different environments: in soils, in air, in foods, etc. (Banke *et al.*, 1997). Chemical and biological investigations on the ethyl acetate extract of *Penicillium* were tested with *Candida albicans* (Petit *et al.*, 2009). *Cladosporium* species are found in soil and on decayed plant material (Revankar and Sutton, 2010).

*Aspergillus* has proven to be a prolific source of secondary metabolites with interesting biological activities, including antibacterial activity. The antibacterial metabolites or extracts were described from terrestrial *Aspergillus* spp. which may have pharmaceutical importance as antibacterial agents (Xu *et al.*, 2015).

The objectives of present research work are to collect the soil samples of two different depths, to isolate fungal strains from soil samples, to investigate antimicrobial activity of fermented broths of all isolated strain and to evaluate antimicrobial activity of all the extracts of isolated strains.

### Materials and Methods

#### Collection of Soil Samples

The two soil samples were collected from the two different depths near Bago River in Dagon Seikkan Township, Ward (94), Yangon Region. The soil samples (10 g for each sample) were taken between twenty centimeter depth and forty centimeter depth and put them into the clean plastic bags. Soil samples were recorded by using the Global Positioning System (GPS) in Table (1). The pH of each sample measured with the pH paper.

- 1) MSc Student, Department of Botany, Dagon University
- 2) Dr, Professor, Department of Botany, Dagon University

Table (1) Location of the collected soil samples

Soil sample	Location	Depth
Sample 1	N 16 ° 49' 34"	Twenty centimeter depth
Sample 2	N 16 ° 49' 34"	Forty centimeter depth

### Composition of culture media (Atlas, 1993)

#### Medium 1

Nutrient Agar Medium (NA)

Nutrient Agar 3.5 g, Distilled Water 100 ml, Agar 1.0 g, pH 6.8

#### Medium 2

Sucrose/Yeast Extract Agar Medium (SYA)

Sucrose 1.0 g, Yeast 0.3 g, Distilled Water 100 ml, Agar 2.5 g, pH 6.8

#### Medium 3

Lactose/Yeast Extract Agar Medium (LYA)

Lactose 1.0 g, Yeast 0.3 g, Distilled Water 100 ml, Agar 2.5 g, pH 6.8

### Antimicrobial Activities of Isolated Fungal Strains

#### Fermentation

The isolated twelve fungal strains grown on 5 days old slant cultures were inoculated into 12 conical flasks (50 ml) containing 20 ml of sucrose/yeast extract medium in each for three day as seed culture. After three days, the seed cultures of twelve strains (1 ml of each) were transferred to twelve fermentation flasks at 30°C, 100 rpm on shaker for 10 days. Inhibitory zones were measured to examine antimicrobial activity of isolated strains (Strobel and Sullivan, 1999).

#### Sucrose/Yeast Extract Medium (SY)

Sucrose 1.0 g, Yeast extract 0.3 g, NaCl 0.3 g, CaCO<sub>3</sub> 0.01 g, Distilled Water 100 ml, pH 6.8

#### Test agar plates

There are five test organisms: three bacterial test organisms (*Agrobacterium tumefaciens*, *Bacillus subtilis* and *Escherichia coli*) and two fungal test organisms (*Candida albicans* and *Malassezia furfur*) in Table (2). Broth culture (0.3 ml) of each test organism is added into 100 ml nutrient agar medium, and then poured into plates.

#### Paper disc diffusion method

After solidification, the paper discs impregnated with fermented broth samples were applied on the test plates. These plates were incubated at 30°C for 24 hr. After 24 to 48 hr, clear zones (inhibitory zones) surrounding the test discs were measured. These zones indicate the presence of the bioactive compounds which inhibit the growth of test organisms (Davis and Stout, 1971).

Table (2) Test organisms and diseases

Test organisms	Diseases
<i>Agrobacterium tumefaciens</i>	Crown gall on plant
<i>Bacillus subtilis</i>	It can cause dysentery, but at the first sign of diarrhoea
<i>Candida albicans</i>	Skin infections, vaginal candidiasis, alimentary tract infections
<i>Escherichia coli</i>	Cholera, diarrhoea and vomiting, urinary tract infection
<i>Malassezia furfur</i>	Dandruff, skin infections

### Antimicrobial activity of crude extracts of fungal strains

The fungal strains were inoculated into the twelve conical flasks containing SY seed medium. Each flask contained 20 ml of medium. After three day, seed cultures (1 ml of each) were transferred to the twelve fermentation flask at 30°C for 7 days on the shaker at 100 rpm. Each fermentation flask contained 20 ml of medium. At day 7 fermentation, the fermented broth of each fungal strain was extracted with ethyl acetate (10 ml) at pH 4.5. Then, the ethyl acetate extracts were dried in the incubator at 40°C. Then, each dried extract was added 0.5 ml of ethyl acetate and mixed thoroughly with glass rod. The extract (20 µl/disc) of each strain was applied for their antimicrobial activity.

## Results

### Isolation of Soil Samples

Twelve fungal strains were isolated from two different soil samples. These strains were given as temporary names HAN 1 to HAN 12 as shown in Table (3) and Fig (1).

Table (3) Isolation of soil fungal strains

Strain	Soil sample	Depth
HAN 1 to HAN 5	Sample 1	Twenty centimeter depth
HAN 6 to HAN 12	Sample 2	Forty centimeter depth



Isolated fungal strains on slant cultures (HAN 1 to HAN 12)

### Antimicrobial activity of fermented broths (day 4 and day 5) of isolated strains

The fermented broths of all strains showed weakly antibacterial activity on *Agrobacterium tumefaciens* at day 4 and day 5 fermentation. All isolated strains showed

weakly antifungal activity on *Malassezia furfur* at day 4 fermentation. Five strains showed weakly antibacterial activity on *Escherichia coli* at day 4 fermentation in Table (4). At day 5 fermentation three strains (HANs 5, 10 & 12) showed moderately antifungal activity but other strains weak activity on *Malassezia furfur*. Eight strains showed weak activity on *E. coli* at day 5 fermentation in Table (5).

Table (4) Inhibitory zones of day 4 fermented broths of isolated strains

Test org. Strain	<i>Agrobacterium tumefaciens</i>	<i>Bacillus subtilis</i>	<i>Candida albicans</i>	<i>Escherichia coli</i>	<i>Malassezia furfur</i>
HAN 1	10	-	-	9	9
HAN 2	10	-	-	-	10
HAN 3	11	-	-	-	10
HAN 4	12	-	-	-	11
HAN 5	11	-	-	-	12
HAN 6	10	-	-	-	10
HAN 7	11	-	-	10	9
HAN 8	10	-	-	-	11
HAN 9	9	-	-	10	9
HAN 10	10	-	-	-	10
HAN 11	10	-	-	9	12
HAN 12	11	-	-	10	12

10-12 mm = weak activity, 13-17 mm = moderate activity, > 18 mm = high activity

Table (5) Inhibitory zones of day 5 fermented broths of isolated strains

	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	10	-	-	10	12
HAN 2	10	-	-	-	11
HAN 3	10	-	-	-	11
HAN 4	9	-	-	10	12
HAN 5	11	-	-	-	13
HAN 6	10	-	-	9	11
HAN 7	9	-	-	9	12
HAN 8	10	-	-	9	12
HAN 9	9	-	-	-	12
HAN 10	10	-	-	10	13
HAN 11	11	-	-	10	11
HAN 12	11	-	-	10	14

#### Antimicrobial activity of fermented broths (day 6 and day 7) of isolated strains

The fermented broths of four strains (HANs 2, 4, 5 & 11) showed moderately activity but other strains weak activity on *A. tumefaciens* at day 6 fermentation. Eleven strains showed weak activity on *E. coli*. At day 6 fermentation, two strains (HANs 1 & 4) showed moderately activity but other strains weak activity on *M. furfur* in Table (6) and Fig. (2). The fermented broths of all twelve strains showed moderately activity on *A. tumefaciens* at day 7 fermentation. Six strains (HANs 1, 2, 6, 9, 10 & 11) indicated better antimicrobial activity than other strains at day 7 fermentation. All strains showed moderate activity on *E. coli*. At day 7 fermentation, six strains (HANs 2, 6, 8, 10, 11 & 12) showed moderately antifungal activity but other strains weak activity on *M. furfur* in Table (7) and Fig. (3).

Table (6) Inhibitory zones of day 6 fermented broths of isolated strains

Strains	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	12	-	-	11	14
HAN 2	13	-	-	9	12
HAN 3	12	-	-	9	11
HAN 4	14	-	-	10	13
HAN 5	13	-	-	9	11
HAN 6	12	-	-	9	12
HAN 7	11	-	-	9	10
HAN 8	10	-	-	9	11
HAN 9	10	-	-	-	9
HAN 10	11	-	-	10	11
HAN 11	14	-	-	11	11
HAN 12	12	-	-	11	10

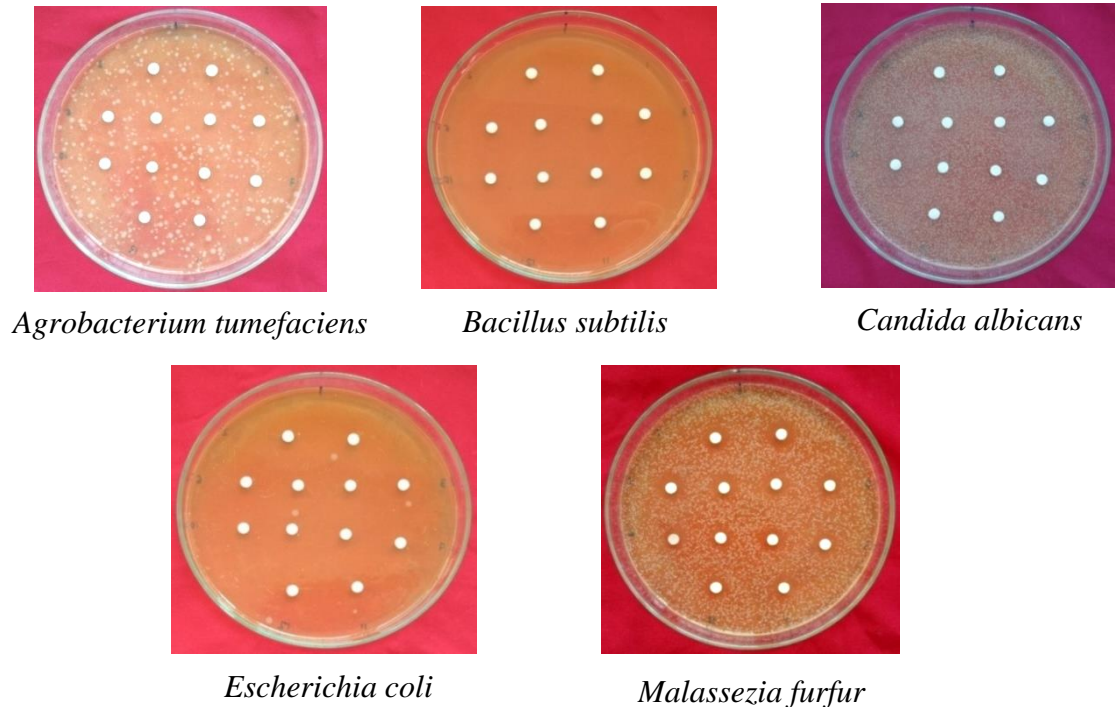


Fig. (2) Inhibitory zones of 6 day fermented broths of isolated twelve strains

Table (7) Inhibitory zones of day 7 fermented broths of isolated strains

Strains	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	15	-	-	13	11
HAN 2	16	-	-	13	13
HAN 3	13	-	-	16	11
HAN 4	14	-	-	16	10
HAN 5	13	-	-	15	12
HAN 6	15	-	-	15	13
HAN 7	14	-	-	16	10
HAN 8	13	-	-	13	13
HAN 9	15	-	-	14	9
HAN 10	15	-	-	15	13
HAN 11	16	-	-	15	13
HAN 12	13	-	-	16	14

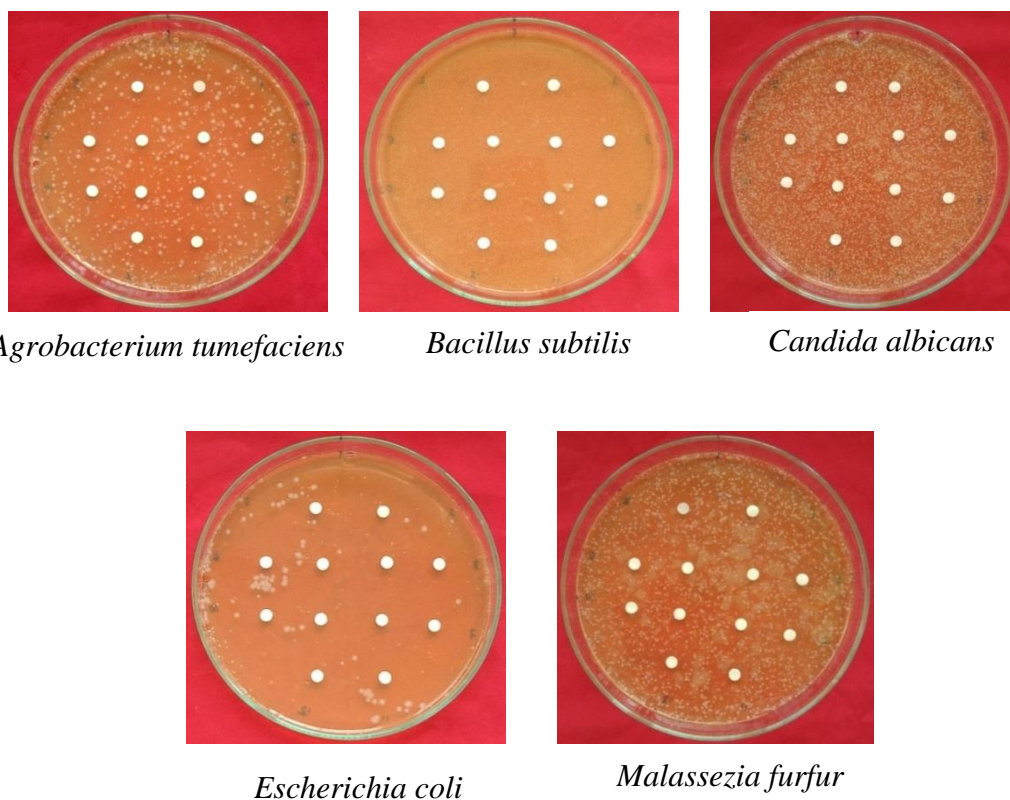


Fig. (3) Inhibitory zones of 7 day fermented broths of isolated twelve strains

#### Antimicrobial activity of fermented broths (day 8 and day 9) of isolated strains

At day 8 fermentation, the fermented broths of five strains (HANs 4, 6, 9, 10 & 11) showed moderately antimicrobial activity but other strains weak activity on *A. tumefaciens* and *M. furfur* and five strains showed moderate activity on *E. coli* in Table (8) and Fig. (4). At day 9 fermentation, two strains (HANs 9 & 10) indicated moderately antibacterial activity but other strains weak activity on *A. tumefaciens*. Seven strains (HANs 1, 2, 3, 4, 5, 6 & 10) indicated moderately antimicrobial activity but other strains weak activity on *M. furfur* in Table (9) and Fig. (5)

Table (8) Inhibitory zones of day 8 fermented broths of isolated strains

Strains	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	11	-	-	12	10
HAN 2	12	-	-	13	12
HAN 3	11	-	-	13	12
HAN 4	13	-	-	15	14
HAN 5	11	-	-	12	11
HAN 6	14	-	-	11	13
HAN 7	12	-	-	14	12
HAN 8	10	-	-	10	11
HAN 9	14	-	-	10	13
HAN 10	15	-	-	12	13
HAN 11	13	-	-	14	15
HAN 12	12	-	-	12	11

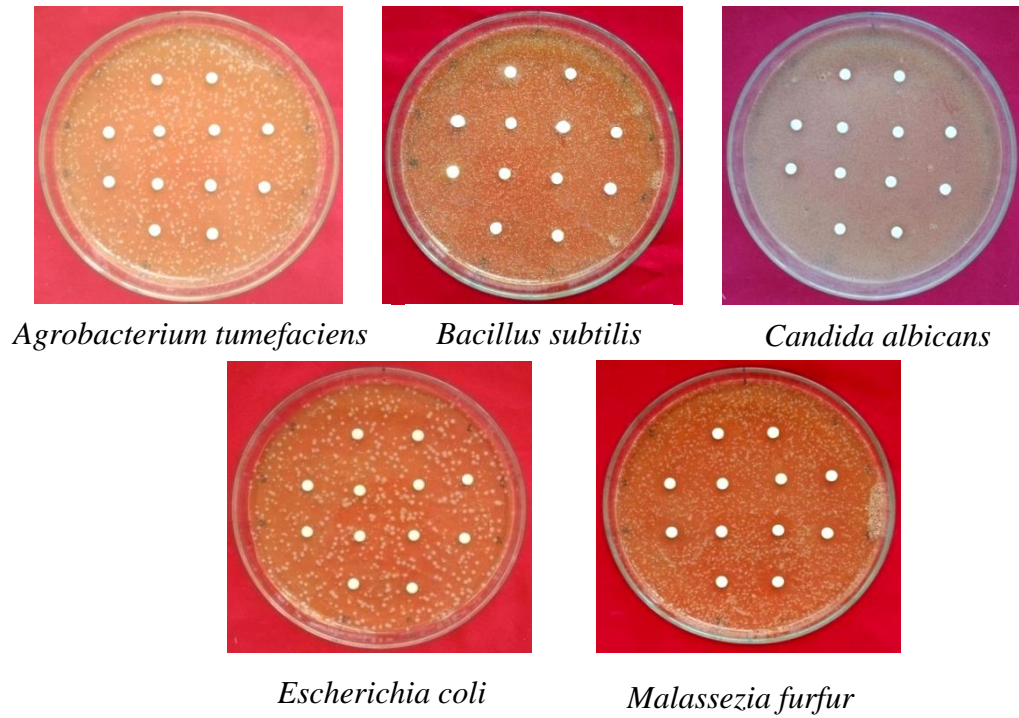


Fig. (4) Inhibitory zones of day 8 fermented broths of isolated twelve strains

Table (9) Inhibitory zones of 9 days fermented broths of isolated strains

Strains	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	11	-	-	11	15
HAN 2	11	-	-	14	16
HAN 3	11	-	-	11	13
HAN 4	12	-	-	12	15
HAN 5	11	-	-	12	13
HAN 6	12	-	-	11	15
HAN 7	11	-	-	12	10
HAN 8	10	-	-	11	10
HAN 9	14	-	-	12	10
HAN 10	14	-	-	12	13
HAN 11	12	-	-	10	10
HAN 12	10	-	-	10	11

10-12 mm = weak activity, 13-17 mm = moderate activity, > 18 mm = high activity



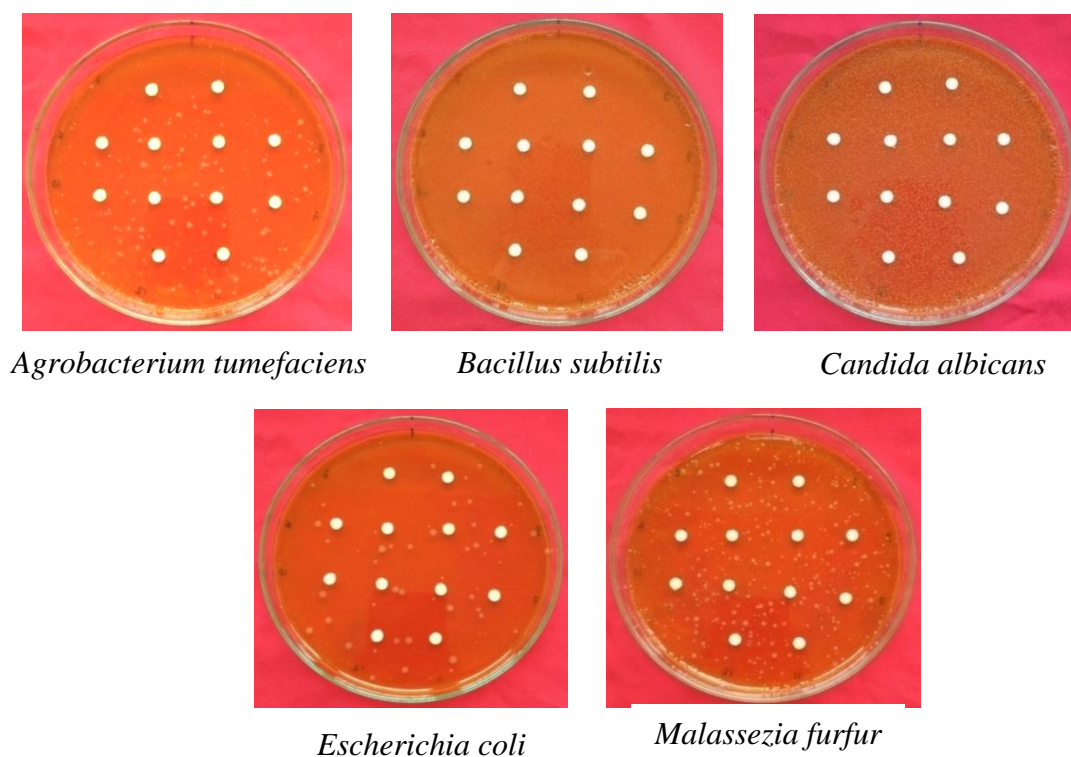


Fig. (5) Inhibitory zones of day 9 fermented broths of isolated twelve strains

#### Antimicrobial Activity of crude extracts of fermented broths

Crude metabolites of twelve strains were extracted with ethyl acetate, and all extracts indicated antimicrobial activity on four test organisms. All crude extracts showed highly antibacterial activity on *A. tumefaciens* (inhibitory zones: 26 mm to 30 mm). All extracts expressed highly antifungal activity on *C. albicans* (inhibitory zones: 22 mm to 29 mm). All extracts indicated highly antibacterial activity on *E. coli* (18 mm to 25 mm). All extracts inhibited highly antifungal activity on *M. furfur* (27 mm to 30 mm) in Table (10) and Fig. (6).

Table (10) Antimicrobial Activity of crude extracts of all strains

Strains	<i>A. tumefaciens</i>	<i>B. subtilis</i>	<i>C. albicans</i>	<i>E. coli</i>	<i>M. furfur</i>
HAN 1	28	-	27	22	30
HAN 2	28	-	29	22	29
HAN 3	30	-	29	22	30
HAN 4	30	-	25	20	27
HAN 5	28	-	25	20	27
HAN 6	30	-	25	24	29
HAN 7	30	-	22	18	28
HAN 8	26	-	22	22	27
HAN 9	26	-	22	22	27
HAN 10	30	-	25	25	30
HAN 11	28	-	25	21	27
HAN 12	28	-	26	24	29

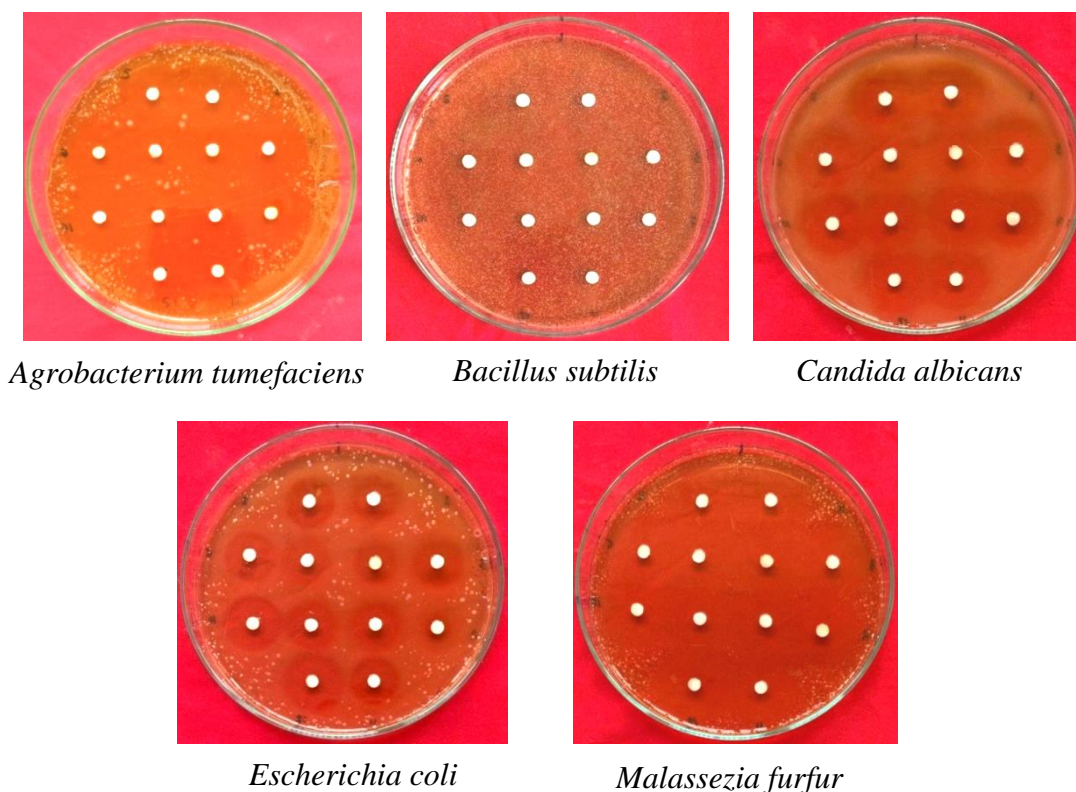


Fig. (6) Antimicrobial activity of crude extracts of isolated strains

### Discussion and Conclusion

In this study, the twelve soil fungi (HAN 1 to HAN 12) were isolated from the two soil samples. Isolated strain HAN 1 was *Cephalosporium* sp., strains HANs 2, 3, 10 were *Aspergillus* spp., strains HANs 4 and 7 were *Penicillium* spp., strain HAN 5 was *Cladosporium* sp., strain HAN 6 was *Madurella* sp., strain HAN 8 was *Paecilomyces* sp., strain HAN 9 was *Hormiscium* sp, strain HAN 11 was *Gonatobotrys* sp. and one strain was unknown. Abraham (2012) has isolated *Cephalosporium* sp. from a sewage outfall in Sardina. Kiroku *et al.*, (1981) has isolated *Cephalosporium gregatum* from bean field soil in Adzuki, they reported antimicrobial activity of this fungus.

In this study, the fermented broths of all isolated strains showed antimicrobial activity on *Agrobacterium tumefaciens* and *Malassezia furfur* from fourth day to ten day fermentation. The crude extracts of twelve strains indicated highly antimicrobial activity on *Agrobacterium tumefaciens*, *Candida albicans*, *Escherichia coli* and *Malassezia furfur*. Yee Yee Thu (2006) has isolated *Aspergillus* species from different soil samples for antimicrobial activity of fermented broth on *C. albicans*, *E. coli* and *M. furfur*. Petit *et al.*, (2009) produced novel antimicrobial secondary metabolites from a *Penicillium* sp. isolated from Brazilian cerrado soil.

Seyedmousavi *et al.*, (2015) stated that *Aspergillus* species are commonly found in soil, decaying vegetation, seeds and grains. Mya Su Maw (2020) isolated fungi from soil samples for antimicrobial activities. Abdulrahman *et al.*, (2022) tested antimicrobial activities (on *C. albicans* and *E. coli*) of metabolites from *Cladosporium* species isolated in Cairo, Egypt. Li *et al.*, (2020) studied secondary metabolites of *Paecilomyces* species, chemical diversity and biological activity such as anticancer activity and antimicrobial activity.

In conclusion, active strains should be continued to produce the bioactive compounds to protect *Agrobacterium tumefaciens* causing crown gall disease on plants, *Candida albicans* causing alimentary tract and vaginal infections, *Escherichia coli*

causing urinary tract infection and *Malassezia furfur* causing dandruff and skin infections on humans. Therefore, active soil fungi were screened and they possessed good antimicrobial activity in this research work so that this research would benefit to produce some antibiotic agents.

#### Acknowledgements

We would like to mention our sincere thanks to Rector, Pro-Rectors from Dagon University, and Professor and Head of Botany Department, DU for their kind permission to submit this paper at 4th Myanmar-Korea Conference on Plant Tissue Culture and Genetics (Useful Plants and Life Science) sponsored by Jeonbuk National University, Korea.

#### References

- Abdulrahman, M.S., Moselhy, S. Mansy, Amal A.I. Mekawey, Hamido M. Hefny and Basma H. Amin. (2022). Antimicrobial Activities of Secondary Metabolites from Airborne *Cladosporium* Species Isolated in Cairo, Egypt.
- Abraham, E.P., (2012). "Cephalosporins 1945-1986". Drugs: 34, 1-14 (1987): SPRINGER LINK.
- Atlas, M. Ronald. (1993). Handbook of Microbiological Media CRC press, London.
- Banke, S.; Frisvad, J.C. and Rosendahl, S. (1997). Taxonomy of *Penicillium chrysogenum* and related xerophilic species, based on isozyme analysis. *Mycol. Res.* 101, 617-624.
- Davis, W.W. and Stout, T. R. (1971). Disc plate method of microbiological antibiotic assay. applied microbiology. Vol.22, No.4.
- Kiroku KOBAYASHI, Fumio TANAKA, Norio KONDO and Tadao UI (1981). A Selective Medium for Isolation of *Cephalosporium gregatum* from Soil and Populations in Adzuki Bean Field Soils Estimated with the Medium, *Ann. Phytopath. Soc. Japan* 47 : 29-34.
- Li, X.Q., Kuo, X., Xin, M. L. and Peng, Z., (2020). A Systematic Review on Secondary Metabolites of *Paecilomyces* species: Chemical Diversity and Biological Activity.
- Lowenfels, J. and Lewis, W. (2006). Teaming with Microbes: A Gardener's Guide to the Soil Food Web, Chapter 3: Bacteria, Timber Press: Portland, Oregon.
- Mendes, R.; Garbeva, P. and Raaijmakers, J.M. (2013). The rhizosphere microbiome: significance of plant beneficial, plant pathogenic and human pathogenic microorganisms. *FEMS Microbiology Reviews*, 35(5):634-s663.
- Mya Su Maw (2020). Isolation of Fungi from Soil Samples and Preliminary Study of Antimicrobial Activities; Maubin University Research Journal 2020, Vol.11.
- Petit, P., Esther M. F. Lucas, Lucas M. Abreu, Ludwig H. P fenning and Jacqueline A. Takahashi (2009). Novel antimicrobial secondary metabolites from a *Penicillium* sp. isolated from Brazilian cerrado soil; *Electronic Journal of Biotechnology* ISSN: 0717-3458; DOI: 10.2225/vol12-issue4-fulltext-9.
- Revankar, S.G., and Sutton, D.A. (2010). Melanized fungi in human disease. *Clin Microbiol Rev.* 23(4): 884-928.
- Seyedmousavi, S., Guillot, J., Arne, P., Hoog, G.S., Mouton, J.W., and Melchers, W.J. (2015). *Aspergillus* and Aspergilloses in wild and domestic animals: a global health concern with parallels to human disease. *Med Mycol.* 2015;53(8):765-97.
- Stroble, R.J. and Sullivan, G.R. (1999). Experimental design for improvement of fermentation, manual of industrial microbiology and Biotechnology, second edition, p 80-102.
- Sylvia, D.M., Hartel, P.G. Fuhrmann, J.J. and Zuberer, D.A. (2005). Principles and Applications of Soil Microbiology (2nd ed.). Edited by David M. Sylvia, Pearson Prentice Hall, Upper Saddle River: New Jersey.
- Xu, W., Li, G., Huang, X. and Luo, Z. (2015). Fungal diversity study in the deep sea sediments of three oceans by culture-dependent approach. *J Appl Oceanogr.* 34:103–110.
- Yee Yee Thu (2006). New Antimicrobial Metabolites Produced by *Trichoderma* sp., *Streptomyces* sp. and *Chaetomium* sp. Isolated from *Mimusops elengi* L., Soil and *Tamarix cananriensis* Willd. PhD Thesis, Department of Botany, University of Yangon.